

# Genomic Analysis of familial Ovarian and Breast Cancer among Saudi Population

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# Acknowledgement

“The authors extend their appreciation to the Deanship of Scientific Research at King Saud University for funding this work through the Undergraduate Research Support Program, Project no. (URSP – 3 –18 – genetic analysis ).”

مطارة البحث العلمي  
DEANSHIP OF SCIENTIFIC RESEARCH

# Introduction

- Breast cancer is the most common cause of mortality of women at the age of 40-50 year old.
- In Saudi women the highest age of diagnosis was between 21 to 36 years which might suggests stronger link to hereditary factors like BRCA mutations.
- Ovarian cancer has the highest mortality rate among gynecological cancers with high morbidity rate estimated to be the seventh most common cancer among women.
- The Saudi population is estimated to have the highest rate of consanguinity worldwide, and the spread of unique genetic homogeneity and Mendelianism has resulted in a higher incidence of genetic disorders.
- There is not any local study that characterized ovarian cancer genetic mutation in Saudi women.

# Introduction

- We aim to identify genetics predisposing factors of familial Ovarian and Breast cancers in Saudi Population.
- Founder mutation identification of breast and ovarian cancers will be of great value to expedite development of screening testing the target population.
- Identification of BRCA mutations in Saudi women will be an important intervention for the patient.
- we aim for (exome) genome sequencing to identify novel mutations in Saudi women and in patient with aggressive clinical features.

# Objectives

- To Screen the patient samples for Epithelial Ovarian and Breast Cancer (excluding borderline tumors) genes which include BRCA1, BRCA2, MLH1, MSH2
- We further want to hypothesize to do whole exome sequencing on a subset of positive samples with evident mutations in BRCA1, BRCA2, MLH1, MLH2. This would enable to find other associated mutation in other physiologically related genes.

# Materials and Methods

## Inclusion Criteria

1. Female patient with confirmed cases epithelial ovarian cancer and breast cancer patients by histopathological examination
2. Saudi nationality at birth, aged 18-80.

## Exclusion Criteria

Known case of BRCA mutation

## Sample storage

Sample type: K3 – EDTA blood Sample

Sample Size: 2 EDTA tubes 3ml Each (one for testing on NGS) and one for confirmation using Sanger sequencing in case a mutation is found.

Sample Storage conditions: there is no limit on the time the sample should be stored, but the sample should be stored at around 4 degrees Celsius (not frozen).

**Transport conditions:** Samples should be transported at 4 degrees Celsius. (The containers arrangement)

# Materials and Methods

## Genetic Screening

Genomic DNA extracted from whole blood is used for genetic analysis. An average of 100-200ng genomic DNA will be utilized for the study. A set of specific mutations in the following genes will be studied –

1. Breast cancer 1 (BRCA1)
2. Breast cancer 2 (BRCA2)
3. mutL homologue 1, colon cancer, nonpolyposis type 2 (MLH1)
4. mutS homologue 2, colon cancer, nonpolyposis type 1 (MSH2)

# Materials and Methods

## DNA extraction and quantification

- The blood samples stored at  $-80^{\circ}\text{C}$  is thawed to room temperature before use.
- The QIAamp DNA Mini kit is used for extraction.
- DNA extraction will be done after lysing the RBCs
- The extracted DNA is tested for quality and quantity on a Nanodrop spectrophotometer.
- The 100 nanogram quantity of DNA is used for PCR amplification.



# Materials and Methods

## Primer design, PCR optimization, amplification and purification

- The primers specific to the polymorphisms are designed using the UCSC Human Genome Browser.
- Primer pairs (forward and reverse) covering a minimum of 50 bases upstream and downstream are sent for commercial synthesis.
- A template of gene sequence with the aligned primers is prepared.
- The PCR program is finalized after several trails by a gradient PCR.
- The optimization is verified by analyzing the amplicon on a 2% agarose gel.

# Materials and Methods

## Primer design, PCR optimization, amplification and purification

- All the samples at 100 ng concentration are used for PCR amplification with Hotstar Taq PCR mastermix from Qiagen.
- The amplification is carried on a Veriti thermal cycler.
- A standard DNA ladder is also run alongside the samples. The separated bands are analyzed on a Biorad Gel Documentation unit.

# Materials and Methods

## Sequence PCR, Purification and DNA Sequencing

- The purified amplified products are used for setting up a sequence PCR. The Big Dye Terminator V3.1 kit from Applied Biosystems is utilized for this purpose.
- Specified volumes of SAM and X terminator solution are added to the sequence amplified products, vortexed at recommended speed for 30 minutes. Samples can be stored at 4°C at this stage for multiplexing on the DNA sequencer later.

## Sequence Analysis and interpretation

A licensed sequence analysis software version 6 is used for the analysis

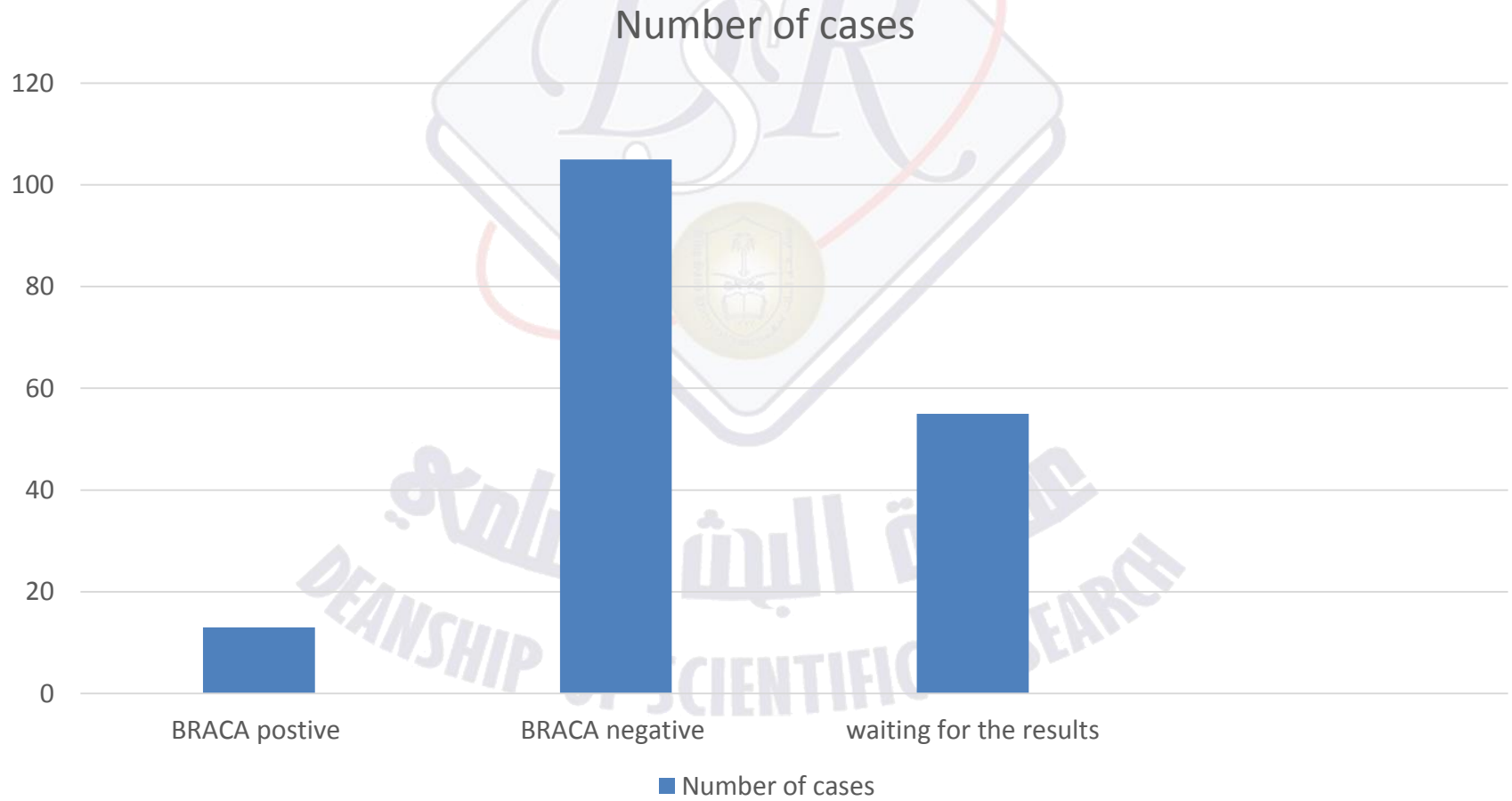
# Results

- The total number of cases collected for BRACA genetic testing are 173 cases during last 2 years (2016-2018)
- 13 cases (7.5 %) was positive for BRACA, 105 (60.6%) was negative and 55 (31.8 %) are waiting for the results.
- Out of the 173 cases, 96 (55.49%) was diagnosed as breast cancer, 8(4.6%) was diagnosed as ovarian-adnexal mass.

# Results

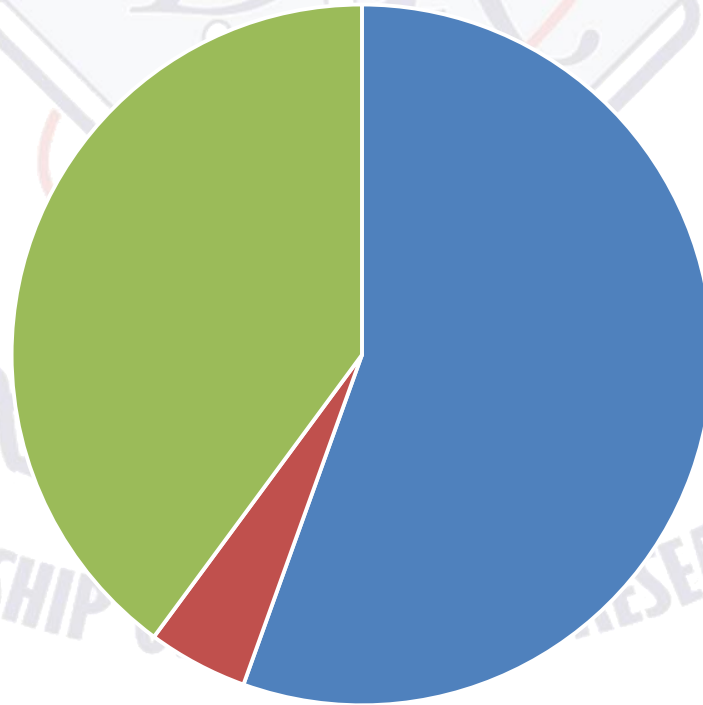
- All of the cases are high risk patient for breast and ovarian cancer, 81 (46.8%) reported a history of breast cancer diagnosed at a young age (premenopausal or young than age 50), 10 (5.8%) has a personal history of triple negative breast cancer diagnosed at age 60 or younger, 6 (3.5%) has personal history of breast cancer affecting both breasts (bilateral breast cancer), 3 (1.73%) has a personal history of both breast and ovarian cancers. 9 (5.2%) had a personal history of ovarian-adnexal cancer.

# Results



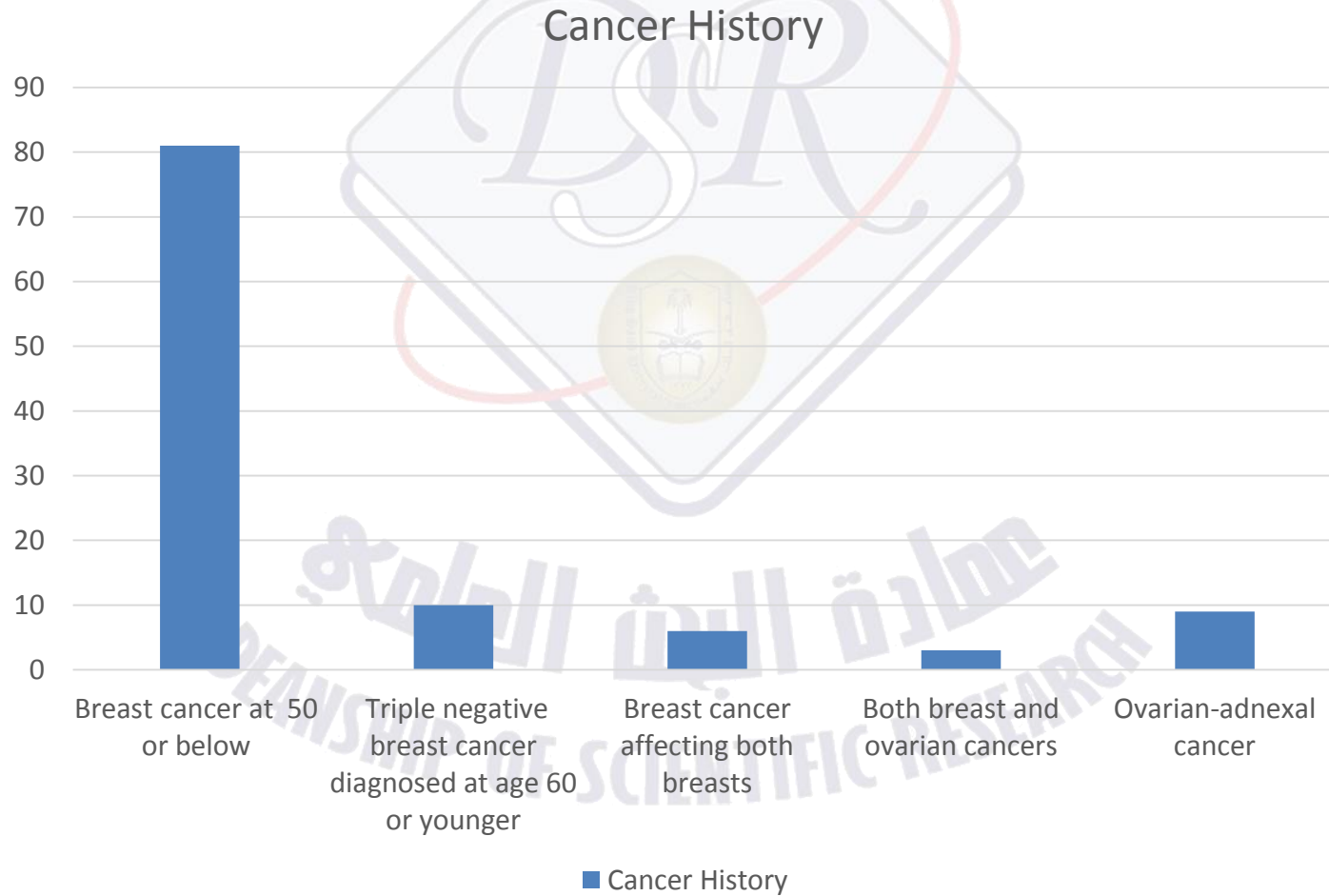
# Results

## Diagnosis of Cancer



■ Breast Cancer ■ ovarian-adnexal mass ■ healthy

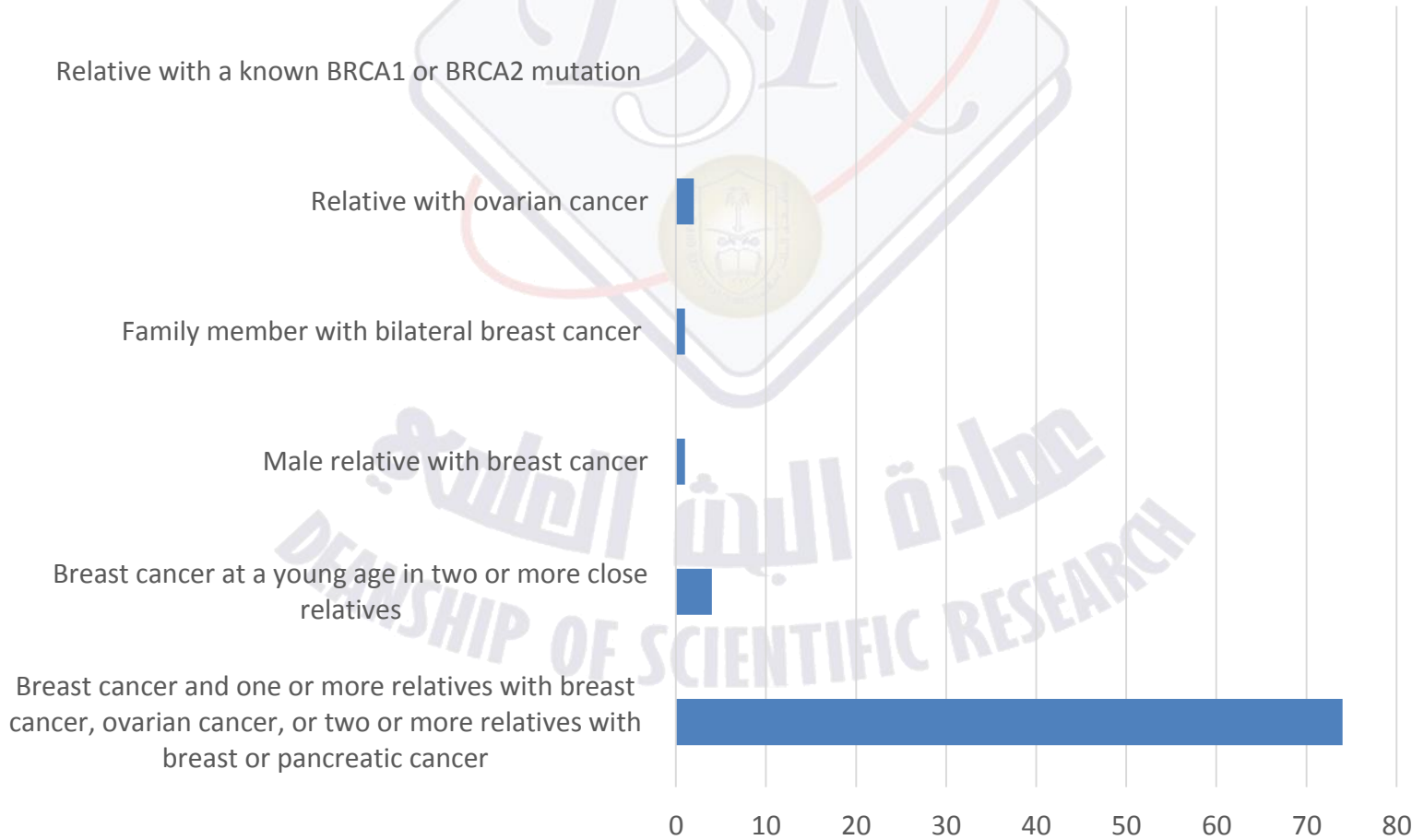
# Results





# Results

## Cancer in Relatives



# Conclusion

- The first phase of the study was concluded, all positive BRACA genes will be further studied for a set of specific genes :
  1. mutL homologue 1, colon cancer, nonpolyposis type 2 (MLH1)
  2. mutS homologue 2, colon cancer, nonpolyposis type 1 (MSH2)
- Whole (exome) genome sequencing will be conducted to identify novel mutations in Saudi women and in patient with aggressive clinical features.

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# References

1. Alghamdi IG, Hussain II, Alghamdi MS, El-Sheemy MA. The incidence rate of female breast cancer in Saudi Arabia: an observational descriptive epidemiological analysis of data from Saudi Cancer Registry 2001-2008. Breast cancer (Dove Med Press [Internet]. Dove Press; 2013 [cited 2017 Jun 8];5:103–9. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/24648763>
2. McPherson K, Steel CM, Dixon JM. ABC of breast diseases. Breast cancer-epidemiology, risk factors, and genetics. BMJ [Internet]. BMJ Publishing Group; 2000 Sep 9 [cited 2017 Jun 8];321(7261):624–8. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/10977847>
3. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin. Jan;61(2):69–90.
4. Registry SNC. Cancer Incidence and Survival Report Saudi Arabia 2007.
5. Jayson GC, Kohn EC, Kitchener HC, Ledermann JA. Ovarian cancer. Lancet [Internet]. 2014 Oct 11 [cited 2017 Jun 8];384(9951):1376–88. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/24767708>
6. Conic I, Dimov I, Tasic-Dimov D, Djordjevic B, Stefanovic V. Ovarian Epithelial Cancer Stem Cells. Sci World J [Internet]. 2011 Jun 9 [cited 2017 Jun 7];11:1243–69. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/21666993>
7. Jelovac D, Armstrong DK. Role of farletuzumab in epithelial ovarian carcinoma. Curr Pharm Des [Internet]. 2012 [cited 2017 Jun 8];18(25):3812–5. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/22591419>